Title

Sickle cell disease: new opportunities and challenges in Africa

Author names

J Makani^{1, 2}, SF Ofori-Acquah³, O Nnodu⁴, A Wonkam^{5, 6}, L Tshilolo⁷, K Ohene–Frempong⁸

Institutional affiliation

- 1. Department of Haematology and Blood Transfusion, Muhimbili University of Health and Allied Sciences, P. O. Box 65001, Dar-es-Salaam, Tanzania
- 2. Nuffield Department of Medicine, University of Oxford
- 3. Children's Healthcare of Atlanta, Center for Endothelial Biology, Emory University, School of Allied Health Sciences, University of Ghana
- 4. Department of Haematology and Blood Transfusion, College of Health Sciences, University of Abuja, Abuja, Nigeria
- 5. Division of Human Genetics, Faculty of Heath Sciences, University of Cape Town South Africa
- 6. Faculty of Medicine and Biomedical Sciences University of Yaoundé I- Cameroon
- 7. Centre Hospitalier Monkole/Cefa, Democratic Republic of Congo
- 8. Children's Hospital of Philadelphia, Philadelphia, USA

Email address

J Makani, julie.makani@muhimbili-wellcome.org Solomon Ofori-Acquah (SOA), <u>soforia@emory.edu</u>, O. Nnodu, <u>oennodu@gmail.com</u> A Wonkam, <u>ambroise.wonkam@uct.ac.za</u> Leon Tshilolo <u>leon.tshilolo2012@gmail.com</u> Kwaku Ohene-Frempong (KOF) ohene@sicklecellghana.org

Abstract

Sickle Cell Disease (SCD) is one of the most common genetic causes of illness and death in the world. This is a review of SCD in Africa, which bears the highest burden of disease. The first section provides an introduction to the molecular basis of SCD and the pathophysiological mechanism of selected clinical events. The second section discusses the epidemiology of the disease (prevalence, morbidity and mortality), at global level and within Africa. The third section discusses the laboratory diagnosis and management of SCD, emphasizing strategies that have proven to be effective in areas with limited resources. Throughout the review, specific activities that require evidence to guide healthcare in Africa, as well as strategic areas for further research will be highlighted.

Introduction

Sickle Cell Disease (SCD) consists of a group of disorders characterised by the presence of sickle haemoglobin. Although over 700 structural hemoglobin (Hb) variants have been identified, only two (Hb S, Hb C) reach high frequencies in Africa. The common SCD syndromes in this region include homozygous HbSS disease (HbSS) commonly known as sickle cell anaemia (SCA) and Hb SC disease. SCD was known in some parts of Africa before the twentieth century: inhabitants of western Africa gave the disease specific names that evoke acute, painful episodes or death, or refer to children destined to die and to be reborn as their own siblings [1],[2]. Africa is the major origin of the sickle (β^{S}) mutations[3]. There are four chromosomal haplotypes that are associated with the β^{S} -mutation. They are named after the regions where they have the highest frequency; Benin, Senegal, Bantu (Central African Region (CAR)) and Arab-Indian. The haplotypes are defined by restriction fragment length polymorphisms (RFLPs) in the β -globin locus (Chen, et al, 1990). Due to the population-specificity of the haplotypes, it is believed that the sickle cell mutation arose independently in these populations and remained to this day[4].

Normal Human Hemoglobin

Human Hb is encoded by a cluster of genes located on chromosomes 11 and 16 that are expressed in a developmentally regulated manner. They are tetramers of two pairs of α -like and β -like globin chains. Adult and fetal hemoglobin have $\alpha \beta$ (Hb A, $\alpha 2\beta 2$), δ (Hb A2, $\alpha 2\delta 2$), or γ chains (Hb F, $\alpha 2\gamma 2$), whereas in the embryo, α -like chains—termed $\zeta \gamma$ (Hb Portland, $\zeta 2\gamma 2$) or $\epsilon \zeta 2\epsilon 2$ — and α and ϵ chains form Hb Gower 2 ($\alpha 2\epsilon 2$) (Figure; [5]).

Embryonic hemoglobin production is confined to the yolk sac. Thereafter the major site of synthesis is the fetal liver. HbF is the predominant type of hemoglobin in fetal life, but around birth there is a switch from fetal to adult globin gene expression, when HbF is gradually replaced by adult hemoglobin, such that by 6 months of age the major Hb is HbA ($\alpha 2\beta 2$). Residual amounts of HbF, however, continue to be synthesized throughout adult life, and the amounts vary considerably, with the majority of adults having less than 1% HbF.



Figure 1 Developmental control of human haemoglobin (Hb) expression [6].

Pathophysiology of clinical events

Sickle haemoglobin (HbS) results from a substitution of one amino acid (Valine) for another amino acid (Glutamic acid) at position six of the β -globin polypeptide chain. This substitution is caused by a single base mutation in codon 6 within the β -globin gene on chromosome 11, where the sequence GAG occurs instead of GTG.

Due to the abnormal amino acid in the β -globin chain, HbS forms long, insoluble polymers when deoxygenated, and the red blood cells (RBCs) containing HbS become less deformable and form a sickle' shape. It was previously thought that the clinical consequences were simply due to this abnormal, rigid sickle red blood cell occluding small blood vessels. However, there is increasing evidence that the pathogenesis of the various clinical events, both acute and chronic, result from a series of complex mechanisms which are

not limited to the RBC. I[7]. These relate to concentration of Hb S and other haemoglobin variants such as HbF within the cell which reduces its ability to polymerise [8], disturbances in the red cell membrane making the cell less responsive to oxidant stress and altered membrane lipids resulting in increased rigidity.[9] [10] [11]. Additionally, adhesion molecules such as integrins ($\alpha_4 \beta_1$), ($\alpha_{\nu} \beta_3$), their receptors (VCAM-1, ICAM-4), selectins interact with endothelial cells, RBC and a variety of soluble proteins within the plasma, such as thrombospondin (from platelets) and von Willebrand factor from endothelial cells to mediate vaso-occlusion within the macro and micro vasculature.[12-16]. Finally there is compelling evidence of the role of nitric oxide (NO) in SCD[17]. NO is a potent regulator of basal vasodilator tone. It also inhibits the expression of cellular adhesion molecules[18]. The increase in haemolysis in SCD results in an excess of haemoglobin in the plasma, which exceeds the scavenging capacity of haptoglobin. The result is that there is abnormal 'cell-free' haemoglobin, which circulates in plasma, binding to and consuming NO, so causing a reduction in the concentration of NO[19]. This results in vasoconstriction, increased adhesiveness of erythrocytes, leukocytes and endothelial cells and platelet aggregation.

Clinical events in SCD

Although SCD stems from an abnormality of the RBC, it is essentially a multi-system disorder, affecting almost every organ-system of the body, as shown in Figure 2. The clinical consequences can be divided into 4 groups; haemolysis and haematological complications, vaso-occlusion, infection and organ dysfunction.



Figure 2 Selected clinical consequences of SCD

Haemolysis and haematological complications

At birth, individuals with SCD do not have anaemia, but with the synthesis of adult Hb, they develop chronic haemolytic anaemia that is present throughout life. This may be interspersed with acute episodes of reduction in haemoglobin – anaemic crises'. Hyperhaemolysis crises is defined by a sudden fall in steady state haemoglobin accompanied by increased reticulocytosis and exaggerated hyperbilirubinaemia. The chronic haemolysis in SCD may result in gall bladder disease due to high levels of bilirubin. Although the main cause of anaemia in SCD is chronic haemolysis, there are other types of anaemia that may occur. Acute splenic sequestration, when there is rapid onset of trapping of red blood cells in the spleen, is characterised clinically by a sudden increase in splenic size, at least 2cm below the left coastal margin, accompanied by a reduction in haemoglobin or haematocrit by 20% of baseline level. This has been described in SCD and is a significant cause of mortality[20]. Anaemia may be secondary to infections such as malaria, bacterial and viral diseases. Of the latter, RBC aplasia in the bone marrow has been notably described and has been associated with infection with parvovirus serotype B19[21].

Vaso-occlusion

Vaso-occlusion (VOC) is thought to be the underlying cause of painful crises, acute splenic sequestration and priapism (painful and prolonged penile erection). Painful crises, considered the hallmark of SCD, are defined as severe pain lasting for 2 or more hours that is attributable to SCD. The sites that are normally affected include the arms, legs, back, abdomen, chest and head. Painful crises do not include other causes/types of pain in SCD such as dactylitis, acute chest syndrome, right upper quadrant syndrome, osteomyelitis and appendicitis. It is the most common cause of hospitalisation and frequent pain (defined as 2 or more painful events a year for three years) is associated with poor quality of life and increased risk of death[22].

Infection

Individuals with SCD are reported to be susceptible to infections with encapsulated organisms such as Streptococcus pneumoniae[23]^{[24],[25]}. The use of oral penicillin in the USA had a significant impact on reduction in mortality[26], and it is now policy in many highincome countries to give penicillin prophylaxis and anti-pneumococcal vaccination to SCD patients[27]. It was previously thought that the situation in Africa may be different. Aside from the fact that the data regarding the clinical spectrum of SCD are limited, there was controversy regarding the role and significance of pneumococcal disease in causing morbidity and mortality in SCD in this setting [28]. However, there is emerging evidence to confirm that pneumococcal disease is a significant cause of bacteraemia in SCD[29], with calls to introduce interventions for preventing infections as a critical factor in improving survival[30, 31]. The various factors that are associated with increased infections in SCD may be directly related or unrelated to the immune system. Some infections may be the result of a complication or treatment of SCD itself. SCD patients are at high-risk of transfusiontransmissible infections particularly with human immunodeficiency virus and viral hepatitis since they receive frequent, often unplanned emergency BT[32-35]. This is particularly important in Africa, given the high prevalence of HIV infection and the operational problems in providing adequate blood-transfusion services. Long-term transfusion may result in iron overload, which in itself is associated with infections due to Yersinia Enterocolitica[36]. SCD causes end-organ damage to the lung, liver, kidney and skin, making these sites susceptible to infection by unusual organisms. In addition, skeletal complications, poor perfusion and blood supply to bone tissue are also thought to contribute to increased susceptibility to infections of the bone, osteomyelitis, which is often due to salmonella infections[37]. Other factors include high bone marrow turnover due to chronic haemolysis which results in increased susceptibility to viral

infection. Parvovirus B19 infections is one of the viral infections that predispose to poor outcome with erythrocytic aplasia that may lead to life-threatening anaemia [21, 38, 39]. However, the epidemiology of this virus in Africa is poorly defined [40-42]. Individuals with SCD may have impairment of the immune system, involving both cellular immunity and humoral immunity. The most well described immune defect is caused by reduced function of the spleen. Patients with SCD have repeated splenic infarction due to vaso-occlusion which causes loss of the splenic vasculature leading to hyposplenism[43]. Reports have suggested that 14% patients with SS-SCD are functionally asplenic at 6 months of age, with this number gradually increasing; 28% at 1 year, 58% at 2 years, 78% at 3 years and by 5 years, 94% are affected[44]. This is from an area without malaria. One of the roles of the spleen is filtration of unopsonised bacteria and remnants of red blood cells from intravascular space as well as opsonised bacteria[45]. Furthermore, the spleen is involved in the synthesis of soluble mediators of immunity. Therefore patients with SCD, with a functional asplenia, have been reported to have impaired antibody responses as well as lacking specific antibodies, particularly against Salmonella species and Streptococcus pneumoniae[46]. This is thought to be due to deficiency of a complement factor involved in the activation of the immune system. The classic pathway is activated by antigen-antibody interaction which causes fixation of complement components C1, C2 and C4 which then activate C3, whereas in the alternate pathway the antigen directly activates C3. Activation of C3, which is an opsonin, results in fixing of antigens on the microorganism[47] making them susceptible to enhanced phagocytosis by neutrophils and monocytes/macrophage. Johnston et al illustrated that patients with SCD have an abnormality in the activation of this pathway with failure of full activation and fixing of C3 to encapsulated bacteria[48]. This results in failure of SCD patients to phagocytose invading organisms, particularly Streptococcus pneumoniae. The distinction between factors directly related to the immune system or not is somewhat arbitrary as there is a lot of overlap between the various factors. Although there have been reports of different patterns of infections in patients with SCD, summarised in table below (Table 1), this review focuses on invasive bacterial infections as detected by blood culture. In the absence of prophylaxis, infections are thought to be the leading cause precipitating clinical events and associated with increased mortality[23, 49].

Syndrome	Organisms	Reference
Septicaemia	S. pneumoniae, H influenza, Salmonella Spp, E	[50],[28],[29]
	Coli, S Aureus, M Pneumoniae	
Pneumonia	S pneumoniae, M Pneumoniae,	
	Chlamydiae	
Meningitis	S. pneumoniae	
Osteomyelitis	Salmonella spp, E Coli, Gram negative organisms,	[51],[37],[52]
	S. Aureus	
Aplastic anaemia	Parvovirus	[21],[38], [39]
AIDS and Hepatitis	HIV Viral hepatitis B,C	[32],[53],[33]
Abdominal pain	Helicobacter pylori, Yersinia enterocolitica	[36]

Table 1 Clinical syndromes and common causative organisms reported in SCD

End-organ dysfunction

With increase in survival, major organs in individuals with SCD are eventually damaged. The brain and lungs are particularly affected, with stroke, defined as an acute neurological syndrome due to vascular occlusion or haemorrhage in which symptoms and signs last for more than 24 hours, being a well-described event. Acute chest syndrome (ACS) is an acute respiratory illness characterised by new pulmonary infiltrates on chest X-ray and falling arterial oxygen saturation [54],[55]. Both these events have been reported to occur with high prevalence in SCD and are also risk factors for death[23, 55, 56].

Heterogeneity of clinical events in SCD

The clinical expression of SCD is heterogeneous. There is inter-individual variability ranging from near complete asymptomatic to severe debilitating illness. There is also variability within an individual, with changes in the type and frequency of clinical events with age. Finally, there is variability in clinical events depending on the geographical location. This is due to the differences in environmental factors such as nutrition, socioeconomic status, and climate that will influence the natural history of disease. The general pattern of clinical disease is characterised by quiescent periods interspersed with acute events, which are referred to as crises.

Clinical event	Prevalence	References
Haemolysis		
Anaemia	Chronic	[57],[58], [59]
Cholelithiasis	Prevalence is 40% by adolescence	[60, 61]
Aplastic anaemia	Associated with parvovirus B19 infection	[62],[61];[63]
Hyperhaemolysis	Limited reports from	[64],[65,
	Africa	661.[67]
Vaso-occlusion		
Pain	More than 60% patients.	[23],[22],[68,
	Most common cause of admission	69]
Acuto splopic	Frequent pain is a risk factor for mortality.	[23 70] [71]
soquestration (ASS)	r requeritiy occurs before the age of Syrs	[23, 70], [71]
Legulcers	Prevalence is 10-25% adults	[72][73]
Priapism	Prevalence is 10-40% males	[74]
	Occurs frequently 5 - 14 years age group	[· ·]
Organ dysfunction		
Neurological events		
Stroke	Prevalence is 10% in	[75]
	children Risk factor for	
	mortality	
	High rate of recurrence	
Cognitive /silent	Prevalence is 20%	[76],[77],[78,
	Risk factor for overt stroke	79]
	Leads to impairment of executive function	
Retinopathy	Prevalence is >30% in HbSC	[80], [80]
Chest		
Acute chest syndrome	Prevalence is 40%.	[54, 55],[56]
(ACS)	Occurs frequently in children,	
	Has severe consequences in	
	adults 12.8 per 100-patient	
Pulmonary hypertension	30%	[81],[82],[83],
	Risk factor for mortality	79, [84]
Avascular necrosis of	Prevalence is 10-50% in adults	[85],[80, 87]
femoral head	Provalance of chronic ranal failure is 5%	1001
Infections	Frevalence of childric terral failure is 5% -	
Malaria		
Bacterial infections	10% children under 5 voors	[03],[30]
	1070 GINULEN UNDER 5 YEARS	[31]

Table 2 The prevalence of selected clinical consequences of SCD

Modified from [92],[93]

The reasons for this heterogeneity are not fully understood[94]. Inter-individual variation in fetal hemoglobin (HbF) levels is one of the main modifiers that contribute to the clinical heterogeneity observed in SCD patients. Higher expression of HbF in adulthood ameliorates morbidity and mortality in SCD[56, 95].

It is now clear that common HbF variation is a quantitative genetic trait shaped by common polymorphisms. Multiple genes, together with an environmental component, determine the measured value of HbF in any given individual. Genetic variation at three major loci accounts for a relatively large proportion (20%–50%) of the phenotypic variation in HbF levels: (1) a singlebase substitution (T/C) at position –158 of the ^G γ globin gene, termed *Xmn*I ^G γ site [96]; (2) the *HMIP* locus (*HBS1L-MYB Intergenic Polymorphism*) on chromosome 6q[97]; and (3) the oncogene *BCL11A* on chromosome 2 [98] These variants have been well reported in nonanemic Northern Europeans and Sardinians, a β -thalassemia cohort, in SCD patients from Brazil, and in the African-American Cooperative Study of Sickle Cell Disease (CSSCD) [99, 100, 101]. There is very little description of the three main genetic polymorphisms explaining phenotypic variation in HbF levels and clinical phenotype in native African SCD patients [97, 102].

Epidemiology of Sickle Cell Disease

Prevalence

The prevalence of SCD can be objectively determined by calculating the birth prevalence of affected children, which requires accurate diagnosis and registration at birth. Since this is not done in most African countries, an alternative method is to use the prevalence of the carrier or heterozygous states (HbAS) to calculate the expected birth rate of SCA based on the gene frequency and Hardy-Weinberg equation. Approximately 300,000 children are born every year with SCD in the world, and countries such as the United States of America, United Kingdom and Jamaica have well-documented SCD population. However, this SCD population constitutes only 1% of the global population of SCD, as over 75% are in Sub-Saharan Africa[103].^[104].It has been estimated that SCD results in the annual loss of several millions of disability-adjusted life years, particularly in the developing world [105]. Hemoglobinopathies alone represent a health burden comparable to that of communicable and other major diseases[106].

Population Genetics and Dynamics: SCD, Malaria and Migration

Compared to non-carriers, healthy carriers of recessive genes for SCD have a well-documented survival advantage against the lethal effects of malaria. As a result, carriers are more likely to reach reproductive age. Consequently, the birth prevalence of SCD is high in Africa [107-109]. The resurgence of malaria in many parts of the world will serve to maintain these polymorphisms, but even if this selective force were removed it would take many generations for the gene frequencies of these conditions to fall significantly[110]. Any changes resulting from variation in selection or population dynamics will, however, be very small compared with the effect of the demographic transition that many countries have undergone over recent vears[110]. Specifically, there is a high prevalence of hemoglobin S (HbS) in Africa and hemoglobin C (HbC) in parts of West Africa[111]. Since subjects that are homozygous for Hb C do not present with severe disease like Hb SS it is anticipated that the frequency of HbC will progressively increase even if malaria is not controlled [112]. Internal migration in Africa has led to SCD, which was previously rare, being introduced in South Africa through an influx of migrants from West and Central Africa[113]. The high birth prevalence of SCD has highlighted the burden of SCD, such that in 2006, the World Health Organization (WHO) recognized SCD as a public health priority[114]. There is limited information about the burden of SCD to the health system and the impact that it has on individuals.

Mortality

There is a higher rate of mortality among individuals with SCD, with reports suggesting that if untreated most children with SCD die in early childhood. Studies done in Nigeria, reported mortality of up to 90%[115] but recent estimates suggest that mortality rate has decreased, and is more likely to be up to 50% by 20 years. This mortality rate in Africa is similar to those reported in the early 1960s in the United States of America and United Kingdom. However, with early diagnosis and comprehensive treatment, significant reductions in mortality have been achieved, with recent reports of improved survival; 85.6% surviving to 18 years in the USA[116], 84% in Jamaica and 99.0% to 16 years in the UK[117]. The common causes of death in the USA, UK and Jamaica are infections, acute splenic sequestration and acute chest syndrome[118], [119], [49], [23], with the highest incidence between 1 and 3 years of age.

Laboratory diagnosis of sickle cell disease

The laboratory diagnosis of SCD is based on the demonstration of HbS and the absence or significant reductions in HbA, with variation in the percentage of two other haemoglobins -

HbF, HbA₂ - in RBCs. Commonly available screening tests in Africa include sodium metabisulphite sickling test and sickle solubility tests, and confirmatory tests using electrophoresis and chromatography to confirm the sickle phenotype (SS/AS/SC/S β ⁻ thalassaemia). The three tests widely used are - haemoglobin electrophoresis, isoelectric focusing (IEF) and High Performance Liquid Chromatography (HPLC). DNA-based assays precisely describe the genotype, however, for clinical purposes, diagnosis usually involves screening (sickling or solubility test) followed by confirmation of the sickle phenotype using gel electrophoresis, IEF or HPLC.

Screening tests

In most African hospitals, screening is done, using the sickling test', which involves making a thin blood film which is then put under hypoxic conditions by the addition of sodium metabisulphite. This will result in RBCs containing HbS becoming deformed (i.e. forming sickle cells) as detected by light microscopy. A "positive" sickling test identifies the presence of sickled RBCs, which occurs in both homo (SS) -and heterozygous (AS) states. The sickle solubility test is another method used for screening which is based on the principle that Sickle Hb becomes insoluble when it is deoxygenated. Additional confirmatory tests are required to confirm SS-SCD or SCD involving other Hb types, when these screening assays are used.

Confirmatory tests

These tests are based on the principle that different haemoglobin isoforms have different overall ionic charge, which makes them migrate with different velocities in an electric field. HBE can be done under alkaline or acidic conditions. HbA, HA₂, HbF and HbS migrate towards the anode under an electric field with different rate of mobility. The results are shown schematically in Figure 8, showing that during alkaline Hb electrophoresis the resolution between HbS and HbF can be poor, particularly in individuals with high HbF levels e.g. neonates. Under acidic conditions, HbF migrates relatively more rapidly and is therefore distinguishable from both HbA and HbS. Isoelectric focusing uses the same principles but is slightly more expensive than HBE. However, it is able to identify more Hb variants that would not be detected by HBE. It also has the advantage that it does not require commercial reagents. HPLC uses cation exchange chromatography to identify the various haemoglobins in an individual. It has the advantage in that it can also accurately quantify the Hb levels. In resource rich countries, screening has largely been replaced by HPLC and confirmation is then done by IEF or HBE. This is mainly because HBE and IEF are labour

intensive, time consuming and would not identify abnormal bands or quantify Hb. Furthermore, the quantification of Hb fractions by HPLC is used to monitor patients who are on Hydroxyurea therapy or exchange blood transfusion.

Molecular diagnosis of SCA

The most popular molecular diagnosis of β^s mutation, based on restriction enzyme digestion, is performed on HBB PCR products. The point mutation, which results in SCD, abolishes the restriction site for the restriction enzyme *Ddel*. Digestion of DNA of individuals homozygous for HbAA, would result in two fragments 188bp and 192bp. Analysis of heterozygous HbAS samples would result in three fragments one of 380bp and the two digested fragments of 180bp and 192 bp. Homozygous HbSS samples would result in 380bp fragments being produced (Figure 4.) This method is simple and cost effective and could be used for prenatal genetic diagnosis in African settings[120].

Figure 3 RFLP of HBB fragment with Ddel. Lane 1: Undigested Control, Lane 2: HbAA control, Lane 3: HbAS control, Lane 4: HbSS MW: Molecular weight marker



Management of Sickle Cell Disease

As a chronic disease, the natural history of SCD is characterised by quiescent periods interspersed by acute events, known as crises, leading to patients seeking health care and frequent hospitalisation. The crises' range from defined syndromes such as acute chest syndrome (ACS), acute splenic sequestration (ASS), to less well defined symptoms that include pain, fever, anaemia, worsening of jaundice and leg ulcers. Other circumstances include pregnancy, dehydration, and extreme cold weather. With the increased life span of individuals with SCD, there has been an increasing awareness of the importance of improving the quality of life as well as preventing damage to major organs. SCD is associated with increased mortality. The causes of mortality in the USA, UK and Jamaica included infections, ACS, ASS, aplastic crises[118], [119], [49], [23]. The management of patients with SCD involves interventions that improve survival, prevent, treat acute events and reduce end-organ damage. Specific conditions or circumstances when SCD patients require extra care are include surgery requiring general anaesthesia, due to increased risk of developing acute sickling complications and sudden death. Over the past 3 decades there has been an improvement in the understanding of the different pathogenic mechanisms responsible for sickle cell events and organ dysfunction. Through a series of clinical trials, effective interventional strategies have been established.

Newborn screening (NBS)

The highest incidence of death occurs in the first 3 years of life[118],[49],[23],[121]. Identification of children at birth by newborn screening (NBS), and institution of preventative care has improved survival[116],[122],[123]. Patients who are identified at birth can be given counselling and advice about the course of illness. They can then be enrolled in comprehensive care programmes that provide prompt and effective care of acute events and prophylaxis against complications, resulting in overall positive impact on survival and quality of life. Countries with large SCD populations and adequate resources have started NBS programmes.

Comprehensive care including dedicated day care facilities

The identification of SCD at birth has to be accompanied by enrolment into programmes that provide comprehensive care by multidisciplinary teams comprising nurses, genetic counsellors, social workers, paediatricians, haematologists, orthopaedic surgeons, ophthalmologists and internists. These programmes provide appropriate advice, counselling

and support to parents and affected individuals. This includes advice such as drinking adequate quantities of fluid to avoid dehydration and wearing warm clothing in cold weather. Specific health education that will enable them to recognise acute events and seek medical care is also essential. Teaching mothers to recognise enlargement of the spleen and anaemia was effective in diagnosing and treating anaemia due to ASS[71],[124]. Patients are also seen on a regular basis and provided with folic acid supplements. The evidence for the burden of folate deficiency in SCD is limited. Prompt treatment of crises (fever and pain), particularly at outpatient or in day-care facilities, has been found to be effective and reduces the burden of hospitalization to the individual and the health system[125],[126],[127], [128]. Long term care should be provided by a multidisciplinary team including professionals who have specialized in haematology and blood transfusion for adults and paediatric haematologists in children. In settings where there is a low prevalence of SCD or limited number of health care professionals, SCD patients can receive care from general health care workers. In such a setting, guidelines for management can be provided to general health care workers with a system of referral to specialised centres.

Prevention and treatment of infections

In the absence of intervention, bacterial infection is the leading cause of mortality in individuals with SCD, and the age group that is most affected is 1 to 3 years [37, 49, 118]. Bacterial infection in SCD is mainly due to Streptococcus pneumoniae, resulting in pneumonia, sepsis and meningitis. The highest incidence of invasive pneumococcal disease is in children less than 6 years of age[118],[91]. In a landmark study in the USA, Gaston and colleagues demonstrated an 84% reduction in incidence of pneumococcal infection with the use of oral penicillin[26]. Interventions with daily oral penicillin and vaccination against pneumococcal infections have successfully reduced mortality in developed countries[26, 116, 129] .In Africa, these interventions have not been implemented as the evidence to demonstrate a similar role of bacterial infections was lacking. This made it difficult for hospitals and governments in developing countries to implement these interventions. Furthermore, published reports have actually questioned the role of prophylaxis against Streptococcus pneumoniae (SPN), in Africa[28]. However, there has been increasing evidence of the role of bacterial infections, particularly due to SPN in causing high childhood mortality[130, 131] Since SCD patients are highly susceptible to SPN infections due to impaired immunity, this makes it even more likely that SPN infections will have a more significant role in SCD mortality. Therefore, there has been an increase in the appeal to implement these interventions[30, 132].

Malaria is widely considered to be one of the major causes of illness and death in patients living with SCD in SSA[90, 104]. Although, SCD individuals have an element of protection against malaria; with a lower prevalence of malaria infection[133-135] and a lower parasite density[136], the risk of mortality when SCD patients get malaria is significantly higher[137]. It is recommended that individuals with SCD who live in a malaria endemic area should receive prophylaxis against malaria[138]. There is ongoing debate as to what is the most appropriate agent that can be used for chemoprophylaxis. The increasing resistance by *Plasmodium falciparum* parasites to Chloroquine has meant that most countries have had to stop using chloroquine. Sulphadoxine-pyrimethamine has anti-folate properties and is not recommended for prophylaxis in patients with SCD who are considered to be folate deficient. Most malaria-endemic countries have therefore been unable to decide which drug to use for prophylaxis in SCD, with options limited to proguanil (paludrine), mefloquine (Lariam), Malarone or Doxycycline. Current practice in malaria-endemic countries involves use of insecticide treated nets and prompt diagnosis and treatment of malaria.

Blood Transfusion (BT)

SCD is contributing to the anaemia in under-fives and pregnant women in areas of high prevalence. Patients with SCD have a compensated chronic haemolytic anaemia which allows them to carry on with normal activities at steady-state haemoglobin with narrow reserve capacity to accommodate strenuous physical activities. The steady state haemoglobin varies from person to person and is related to the level of foetal haemoglobin, co-inheritance of alpha Thalassaemia or heterozygosity for another haemoglobin type such as HbC. Although individuals with SCD have chronic anaemia which is tolerated, rapidly worsening anaemia can occur and this presents as an emergency. It can be caused by ASS, aplastic crises, hyperhaemolysis or associated with other events such as bacterial infections and malaria. Under these circumstances, anaemia is life-threatening and requires prompt treatment with blood transfusion. The products that are used (whole blood or packed RBCs) and the method of transfusion (simple or exchange) are determined by the clinical situation, availability of resources and the capacity to provide the blood product and establish venous access[139]. Blood Transfusion is also effective in other situations, such as acute stroke[140], ACS[141] and peri-operatively[142] Blood Transfusion works by increasing the level of Hb, thus improving oxygen delivery. It also reduces the proportion of sickle RBCs in the circulation. Exchange or red cell transfusion has also been shown to be effective in reducing the level of HbS to less than 30%[143],[144],[145, 146]. This is thought to reduce the deleterious

effects of HbS and improve outcome. Long term blood transfusion therapy (LTBT) has been found to be effective in the prevention of brain injury due to cerebrovascular disease[140]. Blood transfusion is associated with risks which have to be weighed against the benefits when considering implementing this as an intervention. These will be reviewed in the section on stroke.

Pain

Pain, the defining feature of SCD and its commonest symptom, starts early in life and persists throughout life. It is the commonest symptom of SCD and is related to disease severity. Studies in children in developed countries suggest that painful episodes and acute chest syndrome were the most frequent complications of sickle cell disease and that the pain crises is a major predictor of adverse outcome in children along with anaemia and leucocytosis. In adults, large proportion of patients die during an acute episode of pain, making it a risk factor for early death along with acute chest syndrome and stroke. However due to its subjective nature, patients with SCD may not be having appropriate assessment and adequate pain management necessary to prevent complications relating to the pain such as the development of a chronic pain syndrome resulting in worsening of the sickle cell condition. Training is essential for adequate assessment of pain intensity, reporting, documentation by patients, care giver and health workers. Prompt management of pain requires attention to the precipitating causes (stress, infection, dehydration, acidosis, allodynia). Adequate oral analgesic should be administered for mild pain and parenteral for moderate to severe pain according to WHO step ladder for analgesia in patients. When the expected relief is not obtained in response to adequate doses of analgesics, this should alert to the condition of opioid induced hyperaesthesia, allodynia or the progression of acute pain to chronic pain [147-149]. However, many health facilities in Africa do not have access to opiods.

Hydroxyurea

Hydroxyurea (HU) (also known as hydroxycarbamide) has been reported to be effective in improving survival and reducing morbidity in some SCD patients (Table 4). The clinical outcomes include reduction in frequency of painful episodes, and hospital admissions[150]. Hydroxyurea therapy is also monitored by a number of laboratory parameters which include increase HbF levels, mean corpuscular volume (MCV) and reduction in WBC count. Hydroxyurea has been found to be effective in the prevention of brain injury due to cerebrovascular disease[151].

Outcome	Impact in adults	Impact in adolescents
Clinical outcomes		
Pain crises	$\downarrow\downarrow\downarrow\downarrow$	$\downarrow\downarrow$
Hospitalisations	$\downarrow\downarrow\downarrow\downarrow$	$\downarrow\downarrow\downarrow\downarrow$
Blood transfusion therapy	$\downarrow\downarrow\downarrow\downarrow$	\leftrightarrow (insufficient data)
Acute chest syndrome	$\downarrow\downarrow\downarrow\downarrow$	\leftrightarrow (insufficient data)
Laboratory markers		
Foetal haemoglobin	$\uparrow \uparrow \uparrow$	$\uparrow \uparrow \uparrow$
Haemoglobin	$\uparrow \uparrow \uparrow$	\leftrightarrow (not significantly
Mean corpuscular haemoglobin	$\uparrow \uparrow \uparrow$	$\uparrow \uparrow \uparrow$
White blood cell count	$\downarrow\downarrow\downarrow\downarrow$	$\downarrow\downarrow\downarrow\downarrow$
Prevention of end organ damage		
Brain	\leftrightarrow	\leftrightarrow
Spleen	\leftrightarrow	\leftrightarrow
Kidney	\leftrightarrow	\leftrightarrow
Mortality	\downarrow	←→=

Table 3 Summary of study outcomes for hydroxyurea use in adults and children

 $\downarrow\downarrow\downarrow$ =high-grade evidence for decrease; \downarrow =low grade evidence for a decrease; $\uparrow\uparrow\uparrow$ =high-grade evidence for increase; \uparrow =low grade evidence for an increase; \leftrightarrow =not evaluated/not significantly different/insufficient data. Source[152]

Nitric oxide

Lung dysfunction results from a combination of repeated pulmonary infections and infarctions as well as increased vasoconstriction leading to pulmonary hypertension[54]⁻[55]. The latter has recently been shown to be associated with reduced bioavailability of nitric oxide[19], which has resulted in the development of potential therapies such as L-arginine, citrulline and inhaled nitric oxide which is aimed at increasing NO levels through different pathways[153],[154],[155], [156],[157].

Stem cell transplant

The only cure that is available for SCD is stem cell transplantation (SCT), which replaces the host's bone marrow with stem cells containing normal β -globin genotype. Since the first successful transplant reported in 1984[158], there has been significant reduction in risks due to SCT and increasing success, with the best results, of up to 85% event free survival, occurring with HLA-matched sibling donors and transplantation early in the course of the disease before end-organ damage occurs[159]. One limitation of SCT is the availability of sibling donors[160], and therefore there have been attempts to improve survival for unrelated stem-cell donors[161]^{,[162]}. The second limitation of SCT is that this line of treatment requires tremendous

resources, and it becomes increasingly difficult for transplant physicians practicing in the developing world to reconcile the difference between what is possible and what is available. Moreover, it is more difficult to address because the clinical course of SCD is extremely heterogeneous. Despite the knowledge of various genetic and environmental factors known to alter disease severity, it is still difficult to accurately identify children with risk of severe disease before extensive damage has occurred. Until such time that a low-risk, definitive cure is available, the cornerstone of management of SCD is the prevention of early mortality, prevention of end organ damage and improvement of the quality of life.

Gene therapy

Since SCD is caused by a defective gene, definitive treatment would involve replacement of this gene with a normal gene. This has been done successfully in the sickle transgenic mouse[163] but progress in humans has been limited by identification of appropriate vectors and efficacy for gene transfer and low level expression of globin genes.

Role of programmes for control and management of SCD

From a public health perspective, the policy for approaching the control of SCD in national health programmes needs to work in the context of countries with limited resources in health. Although, there is ongoing debate whether care of SCD should be integrated into existing health care services or whether there should be separate disease-specific programmes for SCD, the WHO recommends[164] that for countries where the birth rate of affected infants is above 0.5 per 1,000 births, they should develop separate programmes for these conditions. It is recommended that counties with a high prevalence of SCD, start planning effective rontrol measures. In this context, control of SCD encompasses two elements: providing best possible care for affected individuals and preventing the birth of affected individuals.

With regard to providing best possible care, the following are options, depending on available resources, that have been recommended by Weatherall et al in 2006[105]:

Option one: best possible patient care with the use of prophylactic penicillin following diagnosis, together with retrospective genetic counselling.

Option two: best possible patient care, together with a newborn screening program and the use of penicillin for all homozygous babies, together with retrospective screening and counselling.

Option three: best possible patient care, together with newborn screening and the use of prophylactic penicillin from birth for homozygotes, together with population screening and prospective genetic counselling.

Option four: Option three, plus the availability of prenatal diagnosis, bone marrow transplantation, or both.

The management of SCD involves early diagnosis of affected people, the provision of the most appropriate basic, cost effective treatment and genetic counselling and psychosocial support. The long term goal is to ensure appropriate management at different levels of health care with development of referral centres for specialised diagnosis and treatment. This approach ensures a cost effective way of effectively dealing with a highly prevalent condition in areas where the resources are limited. However, it is important that these centres are not limited to urban areas or centred on academic or research oriented health facilities. In order to avoid this, there must be active strategies to ensure that appropriate management is built into services at all levels of health care with adequate support from these specialised centres. Management of SCD needs to be accompanied by strategies aimed at two levels of prevention: tertiary prevention which involves early diagnosis of SCD and prevention of complications and more ambitiously, secondary prevention which tries to reduce the number of children that are born with SCD. (Note that primary prevention aims to ensure that individuals are born free of SCD). Preventative services involves community education, population screening and genetic counselling that would encourage people to undergo screening before conception, during the antenatal or postnatal period. There are several issues that need to be addressed with regards to prevention of SCD. The aim of screening is to detect SCD in the foetus, discuss the consequences of a diagnosis of SCD and provide options for treatment and prognosis. Since SCD is a recessive disorder, during pre-conception screening, the chances of getting an affected child is variable. There is difficulty in advising a couple not to have children as the risk of getting an affected child may be relatively low (1 in 4), and does not increase with each pregnancy. The highest risk would be for two individuals who are SS who wish to have children. This is different from thalassaemia, where children with the most severe form, thalassaemia major, will inevitably have severe disease. Therefore, one could argue that this therefore justifies the use of pre-natal diagnosis as this would identify pregnancies with SCD children and then parents would be given the appropriate information regarding the consequences and prognosis of SCD and allow more reproductive options to families. Prenatal genetic diagnosis represents one type of

reproductive option as it provides parents with the option to test at-risk pregnancies and make decisions regarding termination of affected pregnancy. The availability and acceptability of prenatal diagnosis and termination of an affected pregnancy are of particular importance in low-resource countries where neither health services, nor families, can afford to pay for longterm treatment of SCA[165]. Close to two-thirds of a sample of 130 Cameroonian parents with affected children reported they would accept termination of an affected pregnancy for SCA [120], a considerably higher proportion when compared to the Cameroonian pre-clinical, clinical medical student and physicians in a previous study (22.4, 10.8 and 36.1%, respectively)[166]. Similar trends were reported in Nigerian parents where 92% of a sample of 53 SCA heterozygous carrier mothers favored prenatal diagnosis, and 63% indicated they would opt for termination of an affected pregnancy[167]. However, in a survey of 403 health workers in a tertiary health care centre in Nigeria, only one third of the respondents accept abortion as an option if prenatal screening is positive for SCA whereas close to half of the respondents (42%). Another study reported that 21.4% Nigerian doctors would accept termination of an affected pregnancy for SCA. [168]. Experience of the effective practice of prenatal genetic diagnosis for SCD (amniocentesis and fetal DNA analysis) was reported in Nigeria and Cameroon[169, 170]. The strong positive views of parents towards prenatal diagnosis and in some cases medical termination of pregnancy may be associated with their experience of affected patients and the psychosocial and/or economic impact of SCA on families. Nevertheless the discrepancy between perception of professional and parents underscores the necessity for more studies to unravel the ethical dilemma around prenatal genetic diagnosis to offer a service that do not conflict with social and cultural values of the affected population. Preimplantation Genetic Diagnosis is a mechanism for accurate genetic diagnosis, careful selection of unaffected embryo and implantation to allow fertile or infertile couples have offspring without SCA. It is an expensive procedure using assisted conception by in vitro fertilization or intracytoplasmic sperm injection. It requires close collaboration between fertility specialists, molecular biologists, geneticists and genetic and fertility counselors and may be more acceptable to individuals of means who may object to prenatal diagnosis followed by termination.

Although SCA is the most severe form of the disease (compared to SC/Sβthalassaemia etc), there is still wide variability in disease severity. Therefore, even with the correct identification and diagnosis of SS with screening, it would be difficult to predict those who would develop severe disease and have a poor outcome.

Conclusion and future challenges

Because of their uneven distribution in high-frequency populations, reflecting their complex population genetics, the true magnitude of burden of SCD is still unknown. In many African countries there are few or virtually no facilities for appropriate diagnosis and management of SCD. There is limited data about frequency, clinical course, or mortality. Without this information it will be impossible to persuade African governments about the burden of this diseaseThe WHO Africa has recommended a set of public health interventions to reduce the burden of SCD in African region namely improving awareness, preventing the disease, early detection, improving the provision of health care for affected individuals by providing effective clinical laboratory, diagnostic and imaging facilities adapted to different levels of the health system, screening of newborns, training of health care workers, developing protocols for treatment, providing genetic counseling, patient support groups, advocacy and research[171]. The situation will be improved by commitment by Member States to integrate SCD prevention and control in national health plans and provide conducive environment for various stakeholders to contribute to the reduction of SCD prevalence, morbidity and mortality. It will also require concerted action on the part of the international community of the richer countries, together with input from other major international health organizations and funding agencies [172, 173].

References

- 1. Onwubalili, J.K., Sickle cell anaemia and reincarnation beliefs in Nigeria. Lancet, 1983. 2(8364): p. 1423.
- 2. Nzewi, E., Malevolent ogbanje: recurrent reincarnation or sickle cell disease? Soc Sci Med, 2001. **52**(9): p. 1403-16.
- 3. Antonarakis, S.E., et al., *Origin of the beta S-globin gene in blacks: the contribution of recurrent mutation or gene conversion or both.* Proc Natl Acad Sci U S A, 1984. **81**(3): p. 853-6.
- 4. Kulozik, A.E., et al., *Geographical survey of beta S-globin gene haplotypes: evidence for an independent Asian origin of the sickle-cell mutation.* Am J Hum Genet, 1986. **39**(2): p. 239-44.
- 5. Weatherall, D.J., *Towards molecular medicine; reminiscences of the haemoglobin field, 1960-2000.* Br J Haematol, 2001. **115**(4): p. 729-38.
- 6. Weatherall, D.J., *Phenotype-genotype relationships in monogenic disease: lessons from the thalassaemias.* Nat Rev Genet, 2001. **2**(4): p. 245-55.
- 7. Stuart, M.J. and R.L. Nagel, Sickle-cell disease. Lancet, 2004. 364(9442): p. 1343-60.
- 8. Nagel, R.L., et al., *Structural bases of the inhibitory effects of hemoglobin F and hemoglobin A2 on the polymerization of hemoglobin S.* Proc Natl Acad Sci U S A, 1979. **76**(2): p. 670-2.
- 9. Noguchi, C.T., D.T. Torchia, and A.N. Schechter, *Intracellular polymerization of Sickle Hemoglobin. Effects of cell heterogeneity*. Journal of Clinical Investigation, 1983. **72**: p. 846-852.
- 10. Brugnara, C., H.F. Bunn, and D.C. Tosteson, *Regulation of erythrocyte cation and water content in sickle cell anemia*. Science, 1986. **232**(4748): p. 388-90.
- 11. Westerman, M.P., et al., *Phase changes in membrane lipids in sickle red cell shed-vesicles and sickle red cells*. Am J Hematol, 1998. **58**(3): p. 177-82.
- 12. Hebbel, R.P., et al., *Erythrocyte adherence to endothelium in sickle-cell anemia. A possible determinant of disease severity.* N Engl J Med, 1980. **302**(18): p. 992-5.
- 13. Frenette, P.S., Sickle Cell Vasoocclusion: Heterotypic, Multicellular Aggregations Driven by Leukocyte Adhesion. Microcirculation, 2004. **11**(2): p. 167-177.
- 14. Okpala, I., Leukocyte adhesion and the pathophysiology of sickle cell disease. Curr Opin Hematol, 2006. 13(1): p. 40-4.
- 15. Ataga, K.I. and E.P. Orringer, *Hypercoagulability in sickle cell disease: a curious paradox.* Am J Med, 2003. **115**(9): p. 721-8.
- 16. Wautier, J.L. and M.P. Wautier, *Erythrocytes and platelet adhesion to endothelium are mediated by specialized molecules*. Clin Hemorheol Microcirc, 2004. **30**(3-4): p. 181-4.
- 17. Villagra, J., et al., *Platelet activation in patients with sickle disease, hemolysis-associated pulmonary hypertension and nitric oxide scavenging by cell-free hemoglobin.* Blood, 2007.
- 18. Gladwin, M.T. and G.J. Kato, *Hemolysis-associated hypercoagulability in sickle cell disease: the plot (and blood) thickens!* Haematologica, 2008. **93**(1): p. 1-3.
- Reiter, C.D., et al., *Cell-free hemoglobin limits nitric oxide bioavailability in sickle-cell disease*. Nat Med, 2002. 8(12): p. 1383-9.
- 20. de Montalembert, M., et al., *Epidemiological and clinical study of sickle cell disease in France, French Guiana and Algeria.* Eur J Haematol, 1993. **51**(3): p. 136-40.
- 21. Pattison, J.R., et al., *Parvovirus infections and hypoplastic crisis in sickle-cell anaemia*. Lancet, 1981. **1**(8221): p. 664-5.
- 22. Platt, O.S., et al., Pain in sickle cell disease. Rates and risk factors. N Engl J Med, 1991. 325(1): p. 11-6.
- 23. Gill, F.M., et al., *Clinical events in the first decade in a cohort of infants with sickle cell disease. Cooperative Study of Sickle Cell Disease.* Blood, 1995. **86**(2): p. 776-83.
- 24. West, T.B., D.W. West, and K. Ohene-Frempong, *The presentation, frequency, and outcome of bacteremia among children with sickle cell disease and fever.* Pediatr Emerg Care, 1994. **10**(3): p. 141-3.
- 25. Wierenga, K.J., et al., *Significance of fever in Jamaican patients with homozygous sickle cell disease.* Arch Dis Child, 2001. **84**(2): p. 156-9.
- Gaston, M.H., et al., Prophylaxis with oral penicillin in children with sickle cell anemia. A randomized trial. N Engl J Med, 1986. 314(25): p. 1593-9.
- 27. American Academy of Pediatrics. Committee on Infectious Diseases. Policy statement: recommendations for the prevention of pneumococcal infections, including the use of pneumococcal conjugate vaccine (Prevnar), pneumococcal polysaccharide vaccine, and antibiotic prophylaxis. Pediatrics, 2000. **106**(2 Pt 1): p. 362-6.
- 28. Kizito, M.E., et al., *Bacteraemia in homozygous sickle cell disease in Africa: is pneumococcal prophylaxis justified?* Arch Dis Child, 2007. **92**(1): p. 21-3.
- 29. Williams, T.N., et al., Bacteraemia in Kenyan children with sickle-cell anaemia: a retrospective cohort and casecontrol study. Lancet, 2009.
- 30. Obaro, S., Pneumococcal Disease in Sickle Cell Disease in Africa: Does Absence of Evidence Imply Evidence of Absence? Arch Dis Child, 2009.
- 31. Tshilolo, L., et al., *Neonatal screening and clinical care programmes for sickle cell disorders in sub-Saharan Africa: lessons from pilot studies.* Public Health, 2008. **122**(9): p. 933-41.

- 32. Bagasra, O., et al., Viral burden and disease progression in HIV-1-infected patients with sickle cell anemia. Am J Hematol, 1998. **59**(3): p. 199-207.
- 33. Diagne, I., et al., [Infections in Senegalese children and adolescents with sickle cell anemia: epidemiological aspects]. Dakar Med, 2000. **45**(1): p. 55-8.
- 34. Hassan, M., et al., *Hepatitis C virus in sickle cell disease*. J Natl Med Assoc, 2003. **95**(10): p. 939-42.
- 35. Tshilolo, L.M., R.K. Mukendi, and S.O. Wembonyama, *Blood transfusion rate in Congolese patients with sickle cell anemia*. Indian J Pediatr, 2007. **74**(8): p. 735-8.
- 36. Blei, F. and D.R. Puder, *Yersinia enterocolitica bacteremia in a chronically transfused patient with sickle cell anemia. Case report and review of the literature.* Am J Pediatr Hematol Oncol, 1993. **15**(4): p. 430-4.
- 37. Barrett-Connor, E., *Bacterial infection and sickle cell anemia. An analysis of 250 infections in 166 patients and a review of the literature.* Medicine (Baltimore), 1971. **50**(2): p. 97-112.
- 38. Serjeant, G.R., et al., *Human parvovirus infection in homozygous sickle cell disease*. Lancet, 1993. **341**(8855): p. 1237-40.
- 39. Smith-Whitley, K., et al., *Epidemiology of human parvovirus B19 in children with sickle cell disease*. Blood, 2004. **103**(2): p. 422-7.
- 40. Jones, P.H., et al., *Human parvovirus infection in children and severe anaemia seen in an area endemic for malaria.* J Trop Med Hyg, 1990. **93**(1): p. 67-70.
- 41. Teuscher, T., B. Baillod, and B.R. Holzer, *Prevalence of human parvovirus B19 in sickle cell disease and healthy controls.* Trop Geogr Med, 1991. **43**(1-2): p. 108-10.
- 42. Yeats, J., H. Daley, and D. Hardie, *Parvovirus B19 infection does not contribute significantly to severe anaemia in children with malaria in Malawi.* Eur J Haematol, 1999. **63**(4): p. 276-7.
- 43. Pearson, H.A., R.P. Spencer, and E.A. Cornelius, *Functional asplenia in sickle-cell anemia*. N Engl J Med, 1969. **281**(17): p. 923-6.
- 44. Brown, A.K., et al., *Reference values and hematologic changes from birth to 5 years in patients with sickle cell disease. Cooperative Study of Sickle Cell Disease.* Arch Pediatr Adolesc Med, 1994. **148**(8): p. 796-804.
- 45. Noel, G.J., S. Katz, and P.J. Edelson, *Complement-mediated early clearance of Haemophilus influenzae type b from blood is independent of serum lytic activity.* J Infect Dis, 1988. **157**(1): p. 85-90.
- 46. Winkelstein, J.A. and R.H. Drachman, *Deficiency of pneumococcal serum opsonizing activity in sickle-cell disease*. N Engl J Med, 1968. **279**(9): p. 459-66.
- 47. Ruddy, S., L.G. Hunsicker, and K.F. Austen, *C3b inactivator of man. 3. Further purification and production of antibody to C3b INA.* J Immunol, 1972. **108**(3): p. 657-64.
- 48. Johnston, R.B., Jr., S.L. Newman, and A.G. Struth, *An abnormality of the alternate pathway of complement activation in sickle-cell disease*. N Engl J Med, 1973. **288**(16): p. 803-8.
- 49. Leikin, S.L., et al., Mortality in children and adolescents with sickle cell disease. Cooperative Study of Sickle Cell Disease. Pediatrics, 1989. 84(3): p. 500-8.
- 50. Okuonghae, H.O., M.U. Nwankwo, and E.C. Offor, *Pattern of bacteraemia in febrile children with sickle cell anaemia*. Ann Trop Paediatr, 1993. **13**(1): p. 55-64.
- 51. Hook, E.W., et al., Salmonella osteomyelitis in patients with sickle-cell anemia. N Engl J Med, 1957. 257(9): p. 403-7.
- 52. Ebong, W.W., Acute osteomyelitis in Nigerians with sickle cell disease. Ann Rheum Dis, 1986. 45(11): p. 911-5.
- 53. Tshilolo, L., R. Mukendi, and R. Girot, *[Sickle cell anemia in the south of Zaire. Study of two series of 251 and 340 patients followed-up 1988-1992]*. Arch Pediatr, 1996. **3**(2): p. 104-11.
- 54. Castro, O., et al., *The acute chest syndrome in sickle cell disease: incidence and risk factors. The Cooperative Study of Sickle Cell Disease.* Blood, 1994. **84**(2): p. 643-9.
- 55. Vichinsky, E.P., et al., *Causes and outcomes of the acute chest syndrome in sickle cell disease. National Acute Chest Syndrome Study Group.* N Engl J Med, 2000. **342**(25): p. 1855-65.
- 56. Platt, O.S., et al., *Mortality in sickle cell disease. Life expectancy and risk factors for early death.* N Engl J Med, 1994. **330**(23): p. 1639-44.
- 57. Hayes, R.J., et al., *The haematology of steady state homozygous sickle cell disease: frequency distributions, variation with age and sex, longitudinal observations.* Br J Haematol, 1985. **59**(2): p. 369-82.
- 58. El-Hazmi, M.A., et al., *The haematological, biochemical and clinical-presentation of haemoglobin S in Saudi Arabia* (*i*). *Haematological & clinical expression*. Trop Geogr Med, 1987. **39**(2): p. 157-62.
- 59. Akenzua, G., et al., *Sickle cell anaemia in Nigeria: a comparison between Benin and Lagos.* Afr J Med Med Sci, 1994. **23**(2): p. 101-7.
- 60. Childs, J.W., Sickle cell disease: the clinical manifestations. J Am Osteopath Assoc, 1995. 95(10): p. 593-8.
- 61. Neonato, M.G., et al., *Acute clinical events in 299 homozygous sickle cell patients living in France. French Study Group on Sickle Cell Disease.* Eur J Haematol, 2000. **65**(3): p. 155-64.
- 62. Serjeant, G.R., et al., *Outbreak of aplastic crises in sickle cell anaemia associated with parvovirus-like agent.* Lancet, 1981. **2**(8247): p. 595-7.
- 63. Juwah, A.I., E.U. Nlemadim, and W. Kaine, *Types of anaemic crises in paediatric patients with sickle cell anaemia seen in Enugu, Nigeria.* Arch Dis Child, 2004. **89**(6): p. 572-6.
- 64. Nolan, V.G., et al., Hemolysis-associated priapism in sickle cell disease. Blood, 2005. 106(9): p. 3264-7.

- 65. Kato, G.J., et al., *Lactate dehydrogenase as a biomarker of hemolysis-associated nitric oxide resistance, priapism, leg ulceration, pulmonary hypertension, and death in patients with sickle cell disease.* Blood, 2006. **107**(6): p. 2279-85.
- 66. Ballas, S.K. and M.J. Marcolina, *Hyperhemolysis during the evolution of uncomplicated acute painful episodes in patients with sickle cell anemia.* Transfusion, 2006. **46**(1): p. 105-10.
- 67. Taylor, J.G.t., et al., *Chronic hyper-hemolysis in sickle cell anemia: association of vascular complications and mortality with less frequent vasoocclusive pain.* PLoS ONE, 2008. **3**(5): p. e2095.
- 68. Olabode, J.O. and W.A. Shokunbi, *Types of crises in sickle cell disease patients presenting at the haematology day care unit (HDCU), University College Hospital (UCH), Ibadan.* West Afr J Med, 2006. **25**(4): p. 284-8.
- 69. Quinn, C.T., et al., *Prognostic significance of early vaso-occlusive complications in children with sickle cell anemia*. Blood, 2007. **109**(1): p. 40-5.
- 70. Topley, J.M., et al., Acute splenic sequestration and hypersplenism in the first five years in homozygous sickle cell disease. Arch Dis Child, 1981. **56**(10): p. 765-9.
- 71. Emond, A.M., et al., *Acute splenic sequestration in homozygous sickle cell disease: natural history and management.* J Pediatr, 1985. **107**(2): p. 201-6.
- 72. Koshy, M., et al., Leg ulcers in patients with sickle cell disease. Blood, 1989. 74(4): p. 1403-8.
- 73. Durosinmi, M.A., S.M. Gevao, and G.J. Esan, *Chronic leg ulcers in sickle cell disease: experience in Ibadan, Nigeria.* Afr J Med Med Sci, 1991. **20**(1): p. 11-4.
- 74. Gbadoe, A.D., et al., [Stuttering priapism in children with sickle cell anemia in Togo.]. Arch Pediatr, 2007. **14**(7): p. 861-863.
- 75. Ohene-Frempong, K., et al., *Cerebrovascular accidents in sickle cell disease: rates and risk factors.* Blood, 1998. **91**(1): p. 288-94.
- 76. DeBaun, M.R., et al., *Cognitive screening examinations for silent cerebral infarcts in sickle cell disease*. Neurology, 1998. **50**(6): p. 1678-82.
- 77. Kinney, T.R., et al., Silent cerebral infarcts in sickle cell anemia: a risk factor analysis. The Cooperative Study of Sickle Cell Disease. Pediatrics, 1999. **103**(3): p. 640-5.
- 78. Miller, S.T., et al., Silent infarction as a risk factor for overt stroke in children with sickle cell anemia: a report from the Cooperative Study of Sickle Cell Disease. J Pediatr, 2001. **139**(3): p. 385-90. t&artType=abs&id=a117580&target=.
- 79. Marouf, R., et al., *Silent brain infarcts in adult Kuwaiti sickle cell disease patients*. Am J Hematol, 2003. **73**(4): p. 240-3.
- 80. Hayes, R.J., P.I. Condon, and G.R. Serjeant, *Haematological factors associated with proliferative retinopathy in sickle cell-haemoglobin C disease*. Br J Ophthalmol, 1981. **65**(10): p. 712-7.
- 81. Castro, O., M. Hoque, and B.D. Brown, *Pulmonary hypertension in sickle cell disease: cardiac catheterization results and survival.* Blood, 2003. **101**(4): p. 1257-1261.
- 82. Gladwin, M.T., et al., *Pulmonary hypertension as a risk factor for death in patients with sickle cell disease*. N Engl J Med, 2004. **350**(9): p. 886-95.
- 83. Ataga, K.I., et al., *Pulmonary hypertension in patients with sickle cell disease: a longitudinal study.* Br J Haematol, 2006. **134**(1): p. 109-15.
- 84. Onyekwere, O.C., et al., *Pulmonary Hypertension in Children and Adolescents with Sickle Cell Disease*. Pediatr Cardiol, 2007.
- 85. Griffiths, J., Avascular necrosis of femoral head in Kenyan africans. East Afr Med J, 1968. 45(9): p. 613-8.
- 86. Ebong, W.W., Avascular necrosis of the femoral head associated with haemoglobinopathy. Trop Geogr Med, 1977. **29**(1): p. 19-23.
- 87. Lee, R.E., J.S. Golding, and G.R. Serjeant, *The radiological features of avascular necrosis of the femoral head in homozygous sickle cell disease*. Clin Radiol, 1981. **32**(2): p. 205-14.
- 88. Abbott, K.C., I.O. Hypolite, and L.Y. Agodoa, *Sickle cell nephropathy at end-stage renal disease in the United States: patient characteristics and survival.* Clin Nephrol, 2002. **58**(1): p. 9-15.
- 89. Fleming, A.F., et al., *Abnormal haemoglobins in the Sudan savanna of Nigeria. I. Prevalence of haemoglobins and relationships between sickle cell trait, malaria and survival.* Ann Trop Med Parasitol, 1979. **73**(2): p. 161-72.
- 90. Fleming, A.F., *The presentation, management and prevention of crisis in sickle cell disease in Africa.* Blood Rev, 1989. **3**(1): p. 18-28.
- 91. Overturf, G.D., D. Powars, and L.J. Baraff, *Bacterial meningitis and septicemia in sickle cell disease*. Am J Dis Child, 1977. **131**(7): p. 784-7.
- 92. Campbell, P.J., et al., *Splenic regrowth in sickle cell anaemia following hypertransfusion*. Br J Haematol, 1997. **96**(1): p. 77-9.
- 93. Yardumian, A. and C. Crawley, Sickle cell disease. Clin Med, 2001. 1(6): p. 441-6.
- 94. Steinberg, M.H., Pathophysiology of sickle cell disease. Baillieres Clin Haematol, 1998. 11(1): p. 163-84.
- 95. Bunn, H.F., Pathogenesis and Treatment of Sickle Cell Disease. N Engl J Med, 1997. 337(11): p. 762-769.
- 96. Labie, D., et al., *Common haplotype dependency of high G gamma-globin gene expression and high Hb F levels in beta-thalassemia and sickle cell anemia patients.* Proc Natl Acad Sci U S A, 1985. **82**(7): p. 2111-4.
- 97. Creary, L.E., et al., *Genetic variation on chromosome 6 influences F cell levels in healthy individuals of African descent and HbF levels in sickle cell patients.* PLoS ONE, 2009. **4**(1): p. e4218.

- 98. Uda, M., et al., Genome-wide association study shows BCL11A associated with persistent fetal hemoglobin and amelioration of the phenotype of beta-thalassemia. Proc Natl Acad Sci U S A, 2008. **105**(5): p. 1620-5.
- 99. Lettre, G., et al., DNA polymorphisms at the BCL11A, HBS1L-MYB, and beta-globin loci associate with fetal hemoglobin levels and pain crises in sickle cell disease. Proc Natl Acad Sci U S A, 2008. **105**(33): p. 11869-74.
- 100. Sedgewick, A.E., et al., *BCL11A is a major HbF quantitative trait locus in three different populations with betahemoglobinopathies.* Blood Cells Mol Dis, 2008. **41**(3): p. 255-8.
- 101. Thein, S.L. and S. Menzel, *Discovering the genetics underlying foetal haemoglobin production in adults.* Br J Haematol, 2009. **145**(4): p. 455-67.
- 102. Makani, J., et al., Genetics of fetal hemoglobin in Tanzanian and British patients with sickle cell anemia. Blood, 2011.
- 103. World Health Organisation. Sickle Cell Anaemia. Agenda item 11.4. in 59th World Health Assembly, 27 May 2006. 2006. Geneva: World Health Organisation.
- 104. Diallo, D. and G. Tchernia, Sickle cell disease in Africa. Curr Opin Hematol, 2002. 9(2): p. 111-6.
- 105. Weatherall, D., et al., *Inherited Disorders of Hemoglobin*, in *Disease Control Priorities in Developing Countries* D. Jamison, Editor. 2006, Oxford University Press: New York. p. 663-680.
- 106. Weatherall, D.J., *Hemoglobinopathies worldwide: present and future*. Curr Mol Med, 2008. 8(7): p. 592-9.
- 107. Enevold, A., et al., *Reduced risk of uncomplicated malaria episodes in children with alpha+-thalassemia in northeastern Tanzania.* Am J Trop Med Hyg, 2008. **78**(5): p. 714-20.
- 108. Williams, T.N., et al., Both heterozygous and homozygous alpha+ thalassemias protect against severe and fatal Plasmodium falciparum malaria on the coast of Kenya. Blood, 2005. **106**(1): p. 368-71.
- 109. Aidoo, M., et al., *Protective effects of the sickle cell gene against malaria morbidity and mortality*. Lancet, 2002. **359**(9314): p. 1311-1312.
- 110. Weatherall, D.J. and J.B. Clegg, *Inherited haemoglobin disorders: an increasing global health problem*. Bull World Health Organ, 2001. **79**(8): p. 704-12.
- 111. Modiano, D., et al., *Haemoglobin S and haemoglobin C: 'quick but costly' versus 'slow but gratis' genetic adaptations to Plasmodium falciparum malaria.* Hum Mol Genet, 2008. **17**(6): p. 789-99.
- 112. Simpore, J., et al., *Modification in the frequency of Hb C and Hb S in Burkina Faso: an influence of migratory fluxes and improvement of patient health care.* Hemoglobin, 2002. **26**(2): p. 113-20.
- 113. Beighton, P. and M.C. Botha, Inherited disorders in the black population of southern Africa. Part I. Historical and demographic background; genetic haematological conditions. S Afr Med J, 1986. **69**(4): p. 247-9.
- 114. World Health Organisation, *Genomics and World Health; Report of the Advisory Committee on Health Research*. 2002, World Health Organisation: Geneva.
- 115. Molineaux, L., et al., Abnormal haemoglobins in the Sudan savanna of Nigeria. III. Malaria, immunoglobulins and antimalarial antibodies in sickle cell disease. Ann Trop Med Parasitol, 1979. **73**(4): p. 301-10.
- 116. Quinn, C.T., Z.R. Rogers, and G.R. Buchanan, *Survival of children with sickle cell disease*. Blood, 2004. **103**(11): p. 4023-4027.
- 117. Telfer, P., et al., *Clinical outcomes in children with sickle cell disease living in England: a neonatal cohort in East London.* Haematologica, 2007. **92**(7): p. 905-12.
- 118. Thomas, A.N., C. Pattison, and G.R. Serjeant, *Causes of death in sickle-cell disease in Jamaica*. Br Med J (Clin Res Ed), 1982. **285**(6342): p. 633-5.
- 119. Brozovic, M. and E. Anionwu, Sickle cell disease in Britain. J Clin Pathol, 1984. 37(12): p. 1321-6.
- 120. Sankaran, V.G. and M.V. Sapp, *Persistence of fetal hemoglobin expression in an older child with trisomy 13.* J Pediatr, 2012. **160**(2): p. 352.
- 121. Lee, A., et al., Improved survival in homozygous sickle cell disease: lessons from a cohort study. BMJ, 1995. **311**(7020): p. 1600-1590.
- 122. Vichinsky, E., et al., Newborn screening for sickle cell disease: effect on mortality. Pediatrics, 1998. 81: p. 749-54.
- 123. Frempong, T. and H.A. Pearson, *Newborn screening coupled with comprehensive follow-up reduced early mortality of sickle cell disease in Connecticut.* Conn Med, 2007. **71**(1): p. 9-12.
- 124. Al-Hawsawi, Z.M. and G.A. Ismail, *Acute splenic sequestration crisis in children with Sickle Cell Disease*. Saudi Med J, 2001. **22**(12): p. 1076-9.
- 125. Wilimas, J.A., et al., A randomized study of outpatient treatment with ceftriaxone for selected febrile children with sickle cell disease. N Engl J Med, 1993. **329**(7): p. 472-6.
- 126. Rahimy, M.C., et al., *Outpatient management of fever in children with sickle cell disease (SCD) in an African setting.* Am J Hematol, 1999. **62**(1): p. 1-6.
- 127. Ware, R.E., S.A. Zimmerman, and W.H. Schultz, *Hydroxyurea as an alternative to blood transfusions for the prevention of recurrent stroke in children with sickle cell disease*. Blood, 1999. **94**(9): p. 3022-6.
- 128. Rahimy, M.C., et al., *Effect of a comprehensive clinical care program on disease course in severely ill children with sickle cell anemia in a sub-Saharan African setting*. Blood, 2003. **102**(3): p. 834-8.
- 129. Knight-Madden, J. and G.R. Serjeant, *Invasive pneumococcal disease in homozygous sickle cell disease: Jamaican experience 1973-1997.* J Pediatr, 2001. **138**(1): p. 65-70.
- 130. Berkley, J.A., et al., *Bacteremia among children admitted to a rural hospital in Kenya*. N Engl J Med, 2005. **352**(1): p. 39-47.

- 131. Roca, A., et al., *Invasive pneumococcal disease in children<5 years of age in rural Mozambique*. Trop Med Int Health, 2006. **11**(9): p. 1422-31.
- 132. de Montalembert, M., V. Brousse, and J.R. Zahar, *Pneumococcal prophylaxis for children with sickle cell disease in Africa.* Arch Dis Child, 2008. **93**(8): p. 715-6.
- 133. Aluoch, J.R., *Higher resistance to Plasmodium falciparum infection in patients with homozygous sickle cell disease in western Kenya*. Trop Med Int Health, 1997. **2**(6): p. 568-71.
- Okuonghae, H.O., M.U. Nwankwo, and E. Offor, *Malarial parasitaemia in febrile children with sickle cell anaemia*. J Trop Pediatr, 1992. 38(2): p. 83-5.
- 135. Kotila, R., A. Okesola, and O. Makanjuola, *Asymptomatic malaria parasitaemia in sickle-cell disease patients: how effective is chemoprophylaxis?* J Vector Borne Dis, 2007. **44**(1): p. 52-5.
- 136. Awotua-Efebo, O., E.A. Alikor, and K.E. Nkanginieme, *Malaria parasite density and splenic status by ultrasonography in stable sickle-cell anaemia (HbSS) children*. Niger J Med, 2004. **13**(1): p. 40-3.
- 137. Makani, J., et al., Malaria in patients with sickle cell anemia: burden, risk factors, and outcome at the outpatient clinic and during hospitalization. Blood, 2010. **115**(2): p. 215-20.
- Oniyangi, O. and A.A. Omari, *Malaria chemoprophylaxis in sickle cell disease*. Cochrane Database Syst Rev, 2006(4): p. CD003489.
- Wahl, S. and K.C. Quirolo, *Current issues in blood transfusion for sickle cell disease*. Curr Opin Pediatr, 2009. 21(1): p. 15-21.
- 140. Adams, R.J., et al., *Prevention of a first stroke by transfusions in children with sickle cell anemia and abnormal results on transcranial Doppler ultrasonography.* N Engl J Med, 1998. **339**(1): p. 5-11.
- 141. Turner, J.M., et al., *Exchange versus simple transfusion for acute chest syndrome in sickle cell anemia adults.* Transfusion, 2009. **49**(5): p. 863-8.
- 142. Vichinsky, E.P., et al., A Comparison of Conservative and Aggressive Transfusion Regimens in the Perioperative Management of Sickle Cell Disease. N Engl J Med, 1995. **333**(4): p. 206-214.
- 143. Stegenga, K.A., et al., *Quality of life among children with sickle cell disease receiving chronic transfusion therapy.* J Pediatr Oncol Nurs, 2004. **21**(4): p. 207-13.
- 144. Prasad, R., et al., *Long-term outcomes in patients with sickle cell disease and frequent vaso-occlusive crises.* Am J Med Sci, 2003. **325**(3): p. 107-9.
- 145. Telen, M.J., Principles and problems of transfusion in sickle cell disease. Semin Hematol, 2001. 38(4): p. 315-23.
- 146. Ohene-Frempong, K., *Indications for red cell transfusion in sickle cell disease*. Semin Hematol, 2001. **38**(1 Suppl 1): p. 5-13.
- 147. Dunlop, R.J. and K.C. Bennett, *Pain management for sickle cell disease*. Cochrane Database Syst Rev, 2006(2): p. CD003350.
- 148. Rees, D.C., et al., *Guidelines for the management of the acute painful crisis in sickle cell disease*. Br J Haematol, 2003. **120**(5): p. 744-52.
- 149. Ballas, S.K., Pain management of sickle cell disease. Hematol Oncol Clin North Am, 2005. 19(5): p. 785-802, v.
- 150. Charache, S., et al., *Effect of hydroxyurea on the frequency of painful crises in sickle cell anemia. Investigators of the Multicenter Study of Hydroxyurea in Sickle Cell Anemia.* N Engl J Med, 1995. **332**(20): p. 1317-22.
- 151. Ware, R.E., M.H. Steinberg, and T.R. Kinney, *Hydroxyurea: an alternative to transfusion therapy for stroke in sickle cell anemia.* Am J Hematol, 1995. **50**(2): p. 140-3.
- 152. National Institutes of Health. National Institutes of Health: Consensus Development Conference Statement: Hydroxyurea Treatment for Sickle Cell Disease. 2008: National Institutes of Health.
- 153. Waugh, W.H., et al., Oral citrulline as arginine precursor may be beneficial in sickle cell disease: early phase two results. J Natl Med Assoc, 2001. **93**(10): p. 363-71.
- 154. Morris, C.R., et al., *Hydroxyurea and arginine therapy: impact on nitric oxide production in sickle cell disease.* J Pediatr Hematol Oncol, 2003. **25**(8): p. 629-34.
- 155. Oppert, M., et al., Inhaled nitric oxide for ARDS due to sickle cell disease. Swiss Med Wkly, 2004. **134**(11-12): p. 165-7.
- 156. Weiner, D.L. and C. Brugnara, *Hydroxyurea and sickle cell disease: a chance for every patient.* Jama, 2003. **289**(13): p. 1692-4.
- 157. Gladwin, M.T., et al., *Nitric oxide donor properties of hydroxyurea in patients with sickle cell disease*. Br J Haematol, 2002. **116**(2): p. 436-44.
- 158. Johnson, F.L., et al., *Bone-marrow transplantation in a patient with sickle-cell anemia.* N Engl J Med, 1984. **311**(12): p. 780-3.
- 159. Walters, M.C., et al., Impact of bone marrow transplantation for symptomatic sickle cell disease: an interim report. Multicenter investigation of bone marrow transplantation for sickle cell disease. Blood, 2000. **95**(6): p. 1918-24.
- 160. Krishnamurti, L., et al., Availability of unrelated donors for hematopoietic stem cell transplantation for hemoglobinopathies. Bone Marrow Transplant, 2003. **31**(7): p. 547-50.
- 161. Woodard, P., B. Lubin, and C.M. Walters, *New approaches to hematopoietic cell transplantation for hematological diseases in children*. Pediatr Clin North Am, 2002. **49**(5): p. 989-1007.
- 162. Adamkiewicz, T.V., et al., *Transplantation of unrelated placental blood cells in children with high-risk sickle cell disease*. Bone Marrow Transplant, 2004.

- 163. Pawliuk, R., et al., Correction of sickle cell disease in transgenic mouse models by gene therapy. Science, 2001. **294**(5550): p. 2368-71.
- 164. World Health Organisation, *Guidelines for the Control of Haemoglobin Disorders (1994)*. 1994, World Health Organisation: Geneva.
- 165. Alwan, A. and B. Modell, *Recommendations for introducing genetics services in developing countries*. Nat Rev Genet, 2003. **4**(1): p. 61-8.
- 166. Wonkam, A., A.K. Njamnshi, and F.F. Angwafo, 3rd, *Knowledge and attitudes concerning medical genetics amongst physicians and medical students in Cameroon (sub-Saharan Africa)*. Genet Med, 2006. **8**(6): p. 331-8.
- 167. Durosinmi, M.A., et al., Acceptability of prenatal diagnosis of sickle cell anaemia (SCA) by female patients and parents of SCA patients in Nigeria. Soc Sci Med, 1995. **41**(3): p. 433-6.
- 168. Adeyemi, A.S. and D.A. Adekanle, *Knowledge and attitude of female health workers towards prenatal diagnosis of sickle cell disease*. Niger J Med, 2007. **16**(3): p. 268-70.
- 169. Akinyanju, O.O., et al., *Initiation of prenatal diagnosis of sickle-cell disorders in Africa*. Prenat Diagn, 1999. **19**(4): p. 299-304.
- 170. Xu, J., et al., *Correction of sickle cell disease in adult mice by interference with fetal hemoglobin silencing.* Science, 2011. **334**(6058): p. 993-6.
- 171. World Health Organisation, Sickle Cell Disease: A Strategy For The WHO Africa Region, R.o.t.R. Director, Editor. 2010.
- 172. Weatherall, D.J., Genomics and global health: time for a reappraisal. Science, 2003. 302(5645): p. 597-9.
- 173. Weatherall, D., et al., *A case for developing North-South partnerships for research in sickle cell disease*. Blood, 2005. **105**(3): p. 921-923.